

Distributional Graphormer (DiG): From Structure Prediction to Distribution Prediction

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Al for Scientific Computation

End-to-end prediction:

Molecular-system descriptor

 → Equilibrium structure, molecular
 property, ...



Protein Structure Prediction

Molecule structure

 → Potential energy (for molecular dynamics simulation)



But the world is *probabilistic* when zooming in:

- Molecular-system descriptor \rightarrow Equilibrium structure r^*
 - **\rightarrow** Structure distribution p(r)



Canonical (const. NVT) https://en.wikipedia.org/wiki/En semble_(mathematical_physics)



AlphaFold cannot predict any of them:



The states of molecules r in real world follow a distribution.



Canonical (const. NVT) https://en.wikipedia.org/wiki/En semble_(mathematical_physics)

Detailed Description:

- In equilibrium: Boltzmann distribution $p(\mathbf{r}) = \exp\left(-\frac{E(\mathbf{r})}{kT}\right)/Z$.
- Non-equilibrium: $q(\mathbf{r})$.
 - $q(\mathbf{r}) = p(\mathbf{z}|\mathbf{x}_1)\delta_{\mathbf{x}_1}(\mathbf{x}),$

where $r \leftrightarrow$ (reaction coord x, other coord z).



The states of molecules r in real world follow a distribution.



Detailed Physics:

- $\begin{array}{c|c} \mathbf{r}^{(1)} \\ \mathbf{r}^{(2)} \end{array} & \text{Macroscopic property: } \mathbb{E}_{q(r)}[f(r)]. \\ \hline \mathbf{r}^{(2)} \end{array} & \text{(Helmholtz) Free energy: } F[q] = \mathbb{E}_{q(r)}[E(r)] Tk\mathbb{E}_{q(r)}[-\log q(r)]. \\ \end{array}$

Entropy S[q]

If
$$q_{x_1}(\mathbf{r}) = q(\mathbf{z}|\mathbf{x}_1)\delta_{x_1}(\mathbf{x})$$
, then
 $F[q_{x_1}] = -kT(\text{ELBO}[q(\mathbf{z}|\mathbf{x}_1), p(\mathbf{r})]$

$$\begin{aligned} q_{\mathbf{x}_1} &= -kT(\text{ELBO}[q(\mathbf{z}|\mathbf{x}_1), p(\mathbf{r})] - \log Z) \\ &= -kT(-\text{KL}(q(\mathbf{z}|\mathbf{x}_1)||p(\mathbf{z}|\mathbf{x}_1)) + \log Z_{\mathbf{x}_1}). \end{aligned}$$

(const. NVT) https://en.wikipedia.org/wiki/En semble_(mathematical_physics)

Canonical

If in partial equilibrium (meta-stable state), $q(\mathbf{z}|\mathbf{x}_1) = p(\mathbf{z}|\mathbf{x}_1)$, then $F[q_{x_1}] = -kT \log \int \exp\left(-\frac{E(x_1, z)}{kT}\right) dz,$ and $p(\mathbf{x}) = \exp\left(-\frac{F[q_x]}{kT}\right)/Z$ is preserved.

The states of molecules *r* in real world follow a distribution.



Canonical (const. NVT) https://en.wikipedia.org/wiki/En semble_(mathematical_physics)

Detailed Physics: Entropy S[q]Macroscopic property: $\mathbb{E}_{q(\mathbf{r})}[f(\mathbf{r})]$. (Helmholtz) Free energy: $F[q] = \mathbb{E}_{q(r)}[E(r)] - Tk\mathbb{E}_{q(r)}[-\log q(r)].$ If $q_{x_1}(r) = p(z|x_1)\delta_{x_1}(x)$, then $\bigwedge F[q_x]$ • $F[q_{x_2}] - F[q_{x_1}]$: without enzyme Reaction free energy activation energy without → stability/concentration. enzyme withenzyme activation • $F[q_{x_3}] - F[q_{x_1}]$: energy with Energy enzyme reactants Activation energy overall energy e.g. C₆H₁₂O₆ + O₂ released during → reaction rate. reaction products CO₂+H₂O \boldsymbol{x}_1 χ_2 \boldsymbol{x}_2

Reaction coordinate https://en.wikipedia.org/wiki/Reaction_coordinate

Querying detailed physics:

- Macroscopic property: $\mathbb{E}_{q(r)}[f(r)]$.
- (Helmholtz) Free energy: $F[q] = \mathbb{E}_{q(r)}[E(r)] Tk\mathbb{E}_{q(r)}[-\log q(r)]$.

Traditional computation:

- $\mathbb{E}_{q(r)}[\cdot]$ or $\mathbb{E}_{p(r)}[\cdot]$: MD/MCMC / umbrella (importance) sampling.
 - \rightarrow convergence issue (gap between state transition time scale (μ s to ms) and affordable simulation time (ps)), auto-correlation / particle degeneracy.

Entropy S[q]

- $\log q(\mathbf{r})$: harmonic approximation / direct free energy estimation.
 - \rightarrow coarse approximation (locally second-order) / need to know reaction coord. x.

Querying detailed physics:

- Macroscopic property: $\mathbb{E}_{q(r)}[f(r)]$.
- (Helmholtz) Free energy: $F[q] = \mathbb{E}_{q(r)}[E(r)] Tk\mathbb{E}_{q(r)}[-\log q(r)]$.

Entropy S[q]

Deep generative models:

- → IID sampling: most sample-efficient way.
- → Capability to approximate the complicated distribution.
- Boltzmann generator [Noé et al., 2019, Science]:
 - → Only applicable to systems with MD data.
- Distributional Graphormer (DiG):
 - → Transferable to a range of systems.



















Structure generation

Density calculation

Conditional generation

Capabilities



Equivariant Graphormer

Invariant distribution = invariant prior + equivariant score model.

Graphormer to encode complex structure
 Minimum inductive bias to ensure



Minimum inductive bias to ensure equivariance



Challenges in Model Training

Data scarcity

Need stepwise training signals

- Hard to collect sufficient experimental or simulation data to well characterize the equilibrium distribution for various systems.
- Supervision is only available at the end of the diffusion process.
- Backprop through the whole diffusion-process simulation is very costly.

Training from Energy Function & Simulation Data

• Physically-Informed Pre-training

$$\sum_{m} \left\| \mathbf{s}_{\theta,0} \left(\mathbf{R}^{(m)} \right) + \nabla E \left(\mathbf{R}^{(m)} \right) \right\|^{2} + \sum_{t} \sum_{m} \left\| \frac{\beta_{t}}{2} \nabla \left(\left(\mathbf{R}^{(m)}_{t} + \nabla \right) \cdot \mathbf{s}_{\theta,t} \left(\mathbf{R}^{(m)}_{t} \right) + \left\| \mathbf{s}_{\theta,t} \left(\mathbf{R}^{(m)}_{t} \right) \right\|^{2} \right) - \partial_{t} \mathbf{s}_{\theta,t} \left(\mathbf{R}^{(m)}_{t} \right) \right\|^{2}.$$

Energy-function supervision

- Propagate the supervision by consistency with Fokker-Planck equation
- Stepwise loss function
- Training with Simulation Data

Data supervision

 $\sum_{t} \sum_{n} \mathbb{E}_{\boldsymbol{\epsilon} \sim \mathcal{N}(0,\mathbf{I})} \| \sigma_{t}^{2} \mathbf{s}_{\theta,t} (\alpha_{t} \mathbf{R}^{(n)} + \sigma_{t} \boldsymbol{\epsilon}) + \boldsymbol{\epsilon} \|^{2}.$

Protein Conformation Sampling

SARS-CoV-2 main protease (PDB id: 6lu7, 306 residues)

- Ground-truth: MD Simulation from Folding@home, 2.6 ms. Est. 26k GPU days.
- DiG sampled structures: ~40k structures, ~18 GPU days.

structures in whole functional relev

Alphafold <u>only</u> gradin Similar to this one.



Protein Conformation Sampling

SARS-CoV-2 spike receptor-binding domain (PDB id: 6m0j, 229 residues)

- Ground-truth: MD Simulation from Folding@home, 1.8 ms. Est. 20k GPU days.
- EDP sampled structures: ~80k structures, ~18 GPU days.





Sampling Meta-Stable Structures

■ Adenylate kinase 腺苷酸激酶



4ake

- 1ake
- Human B-Raf kinase 人类 B-Raf 激酶



■ LmrP membrane protein LmrP 膜蛋白



D-Ribose binding protein

大肠杆菌 D-核糖结合蛋白



2dri

1urp

Conformation Transition Pathway Prediction





Adenylate Kinase (open ↔ close) 腺苷酸激酶

LmrP membrane protein (open ↔ close) LmrP 膜蛋白

Protein-Ligand Binding Sampling



与红细胞和血小板的生成相关,在一些免疫过程中起关键作用



p38 丝裂原活化蛋白激酶

参与细胞生理和病理过程,包括凋亡、应激、进入细胞周期、炎症反应

Catalyst Adsorption Sampling

Highlights:

- Generalizable to unseen systems.
- Discovered new adsorption sites (verified by DFT).
- Speed-up:
 - Classical (DFT MD/Relaxation): days.
 - DiG: seconds.







CH3-C-O- on Ti-Ir surface

Density Estimation

single N or O atom on **C**.



By DiG

By DFT (reference)





























Inverse Design

Band Gap = 0 eV → **Graphite**

Band Gap = 4 eV → **Diamond**





